

Visually Assessing Maize Leaves: From Spectral Sampling to High-Fidelity Color Reproduction

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Outline

- Introduction
- Methodology
- Results and Discussion
- Conclusion

Introduction

➤ Context

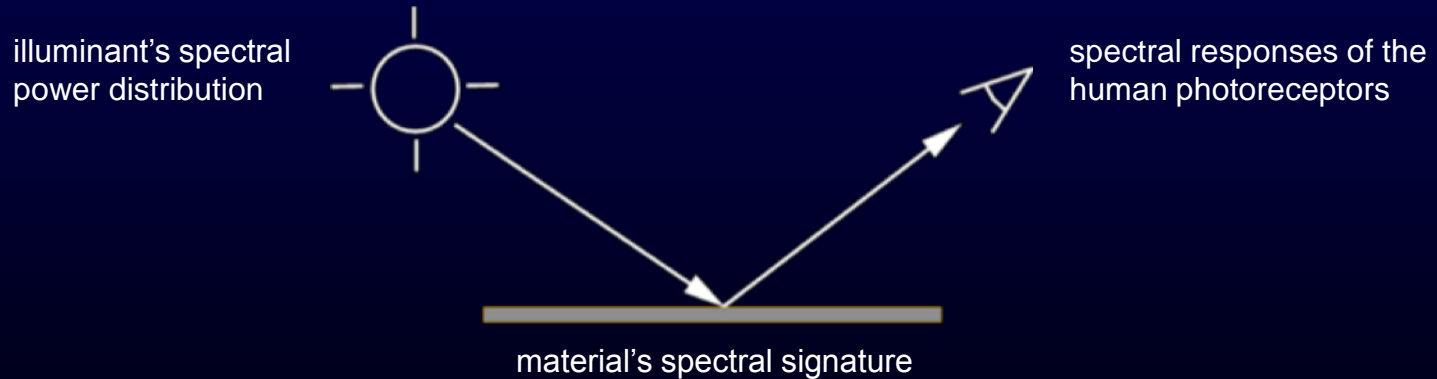
- Maize (corn) crops are extensively used in food and biofuel production worldwide



- A number of protocols have been proposed to use leaf color as an indicator of biophysical phenomena affecting these plants' physiology and appearance
- Several vegetation indices, employing leaf's reflectance and transmittance samples, have also been formulated to monitor the development of these plants

➤ Pros and cons of each strategy

- The complexity of the color perception process can make the correct interpretation of a leaf's chromatic attributes a difficult task



- Spectral vegetation indices:
 - are not subject to color perception issues (e.g., metamerisms)
 - require a number of spectral samples obtained using specialized sensors
 - are usually formulated to assess specific conditions affecting the plants
- The visual feedback provided by foliar chromatic attributes allows for a rapid screening of the net effect of several factors affecting a plant

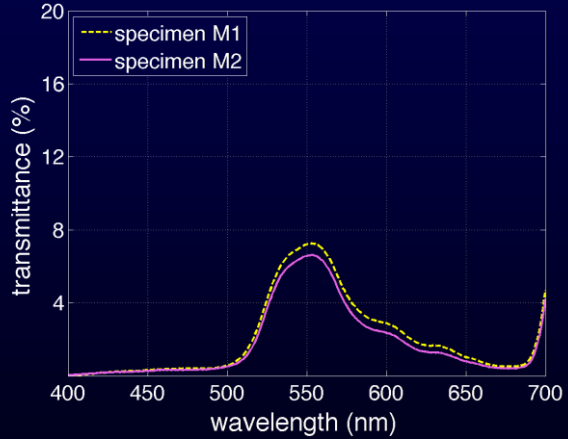
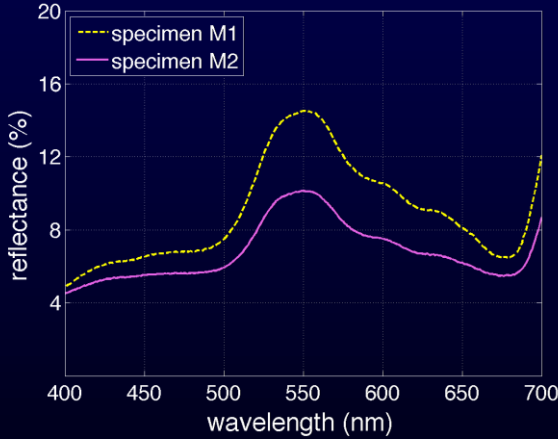
- How about the combined use of indices and color-based strategies?
 - Leaf chromatic attributes can be obtained using spectral reflectance and transmittance samples employed in the computation of vegetation indices
 - Ideally, one would like to employ a number of spectral samples that would maximize the color fidelity to sensor costs ratio

- How many spectral reflectance and transmittance samples would be sufficient to obtain a high-fidelity reproduction of maize leaves' colors?

Methodology

➤ Materials

- Specimens (M1 and M2) with measured spectral reflectance and transmittance curves made available in the LOPEX (Leaf Optical Experiments 1993) database



angle of incidence of 8 degrees

➤ Approach

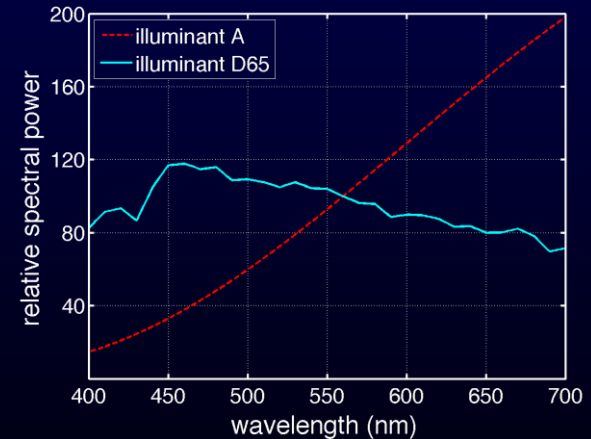
- Computation of maize leaves' chromatic attributes considering distinct light interaction scenarios and sparse spectral sampling resolutions



- Visual inspection of colored swatches (generated using the computed attributes)
- Assessment of their color fidelity using a device independent CIE-based metric

➤ Light interaction variables

- Specimens' light propagation behaviours
 - Reflected light only (e.g., leaf over an opaque surface)
 - Reflected and transmitted light
- Indoors and outdoors illumination settings
 - Standard CIE A illuminant (tungsten lamp)
 - Standard CIE D65 illuminant (average daylight)



➤ Selected spectral resolutions

- Illuminants' spectral power curves and specimens' spectral reflectance and transmittance curves are sampled using the same spectral intervals

N	Spectral Intervals	Sampled Wavelengths
3	Variable	465, 551 and 608 <i>nm</i> (monitor chromaticities)
5	75 <i>nm</i>	400, 475, 550, 625 and 700 <i>nm</i>
6	60 <i>nm</i>	400, 460, 520, 580, 640 and 700 <i>nm</i>
7	50 <i>nm</i>	400, 450, 500, 550, 600, 650 and 700 <i>nm</i>
8	37 <i>nm</i>	400, 437, 474, 511, 548, 585, 622, 659 and 696 <i>nm</i>
301	1 <i>nm</i>	all from 400 to 700 <i>nm</i> (full spectral resolution)

- Standard monitor chromaticity coordinates are used for comparison purposes
- Full spectral resolution ($N=301$) employed as a reference for fidelity assessments

➤ CIELAB differences between pairs of swatches computed as:

$$\Delta E_{ab}^* = \sqrt{(L_s^* - L_f^*)^2 + (a_s^* - a_f^*)^2 + (b_s^* - b_f^*)^2},$$

where:

L^* , a^* and b^* are CIELAB color space dimensions, and the subscripts s and f represent the sparse and full spectral resolutions, respectively.

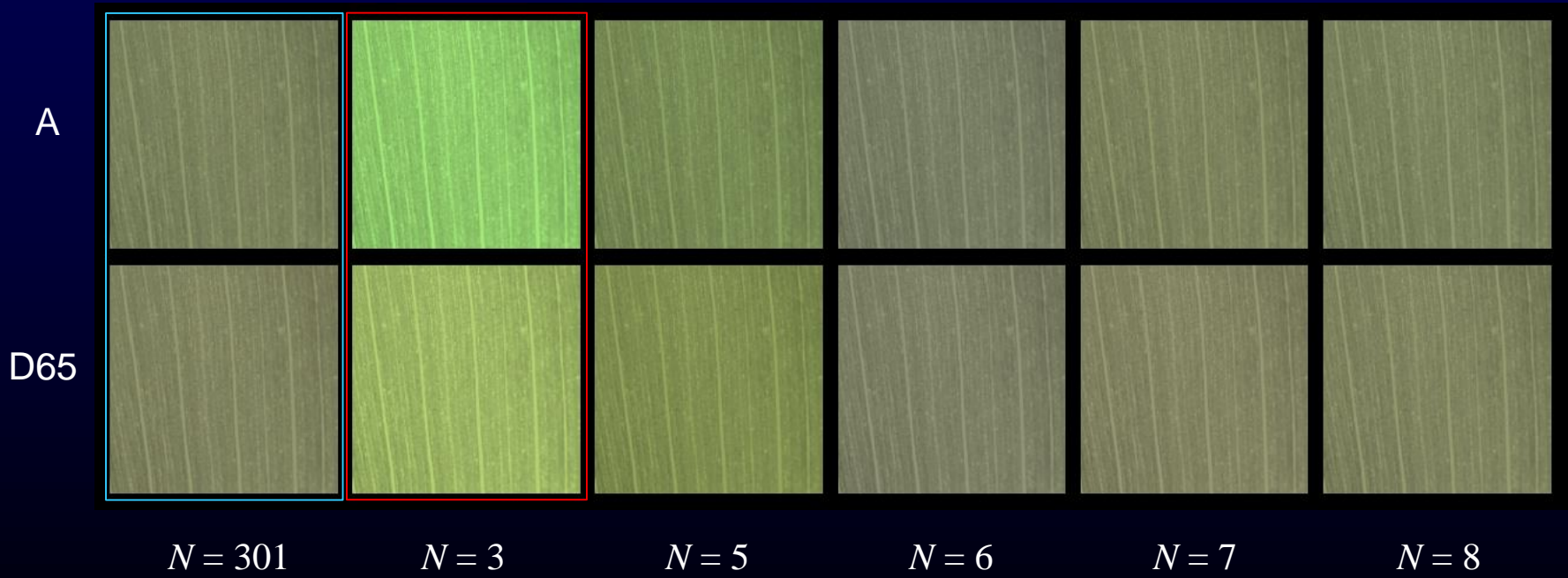
Perceptibility threshold: $\Delta E_{ab}^* < 2.3$

(Source: Mahy *et al.*, 1994)

Results and Discussion

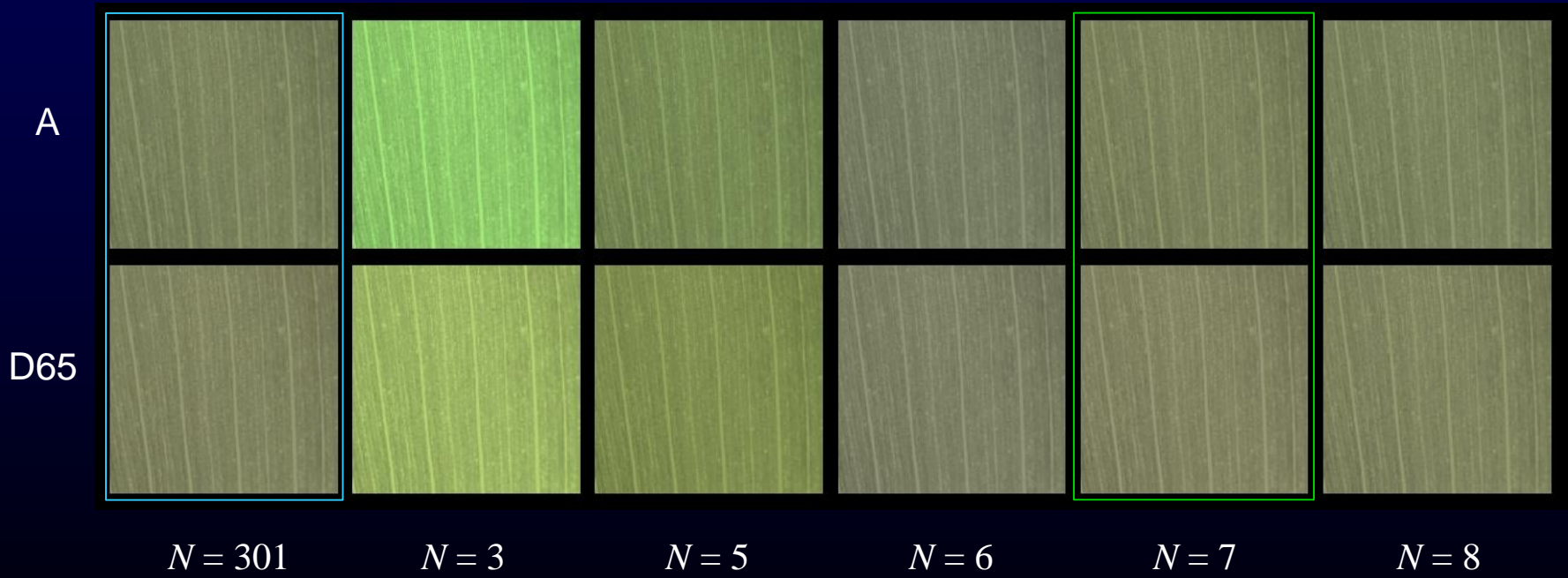
➤ Specimen M1

- Swatches depicting foliar reflective behaviour



➤ Specimen M1

- Swatches depicting foliar reflective behaviour



- CIELAB differences

Illuminant	Reflective Behaviour				
	$N = 3$	$N = 5$	$N = 6$	$N = 7$	$N = 8$
A	46.7559	11.2375	6.4528	0.8461	0.8020
D65	36.1066	19.7921	5.5571	2.0533	1.3456

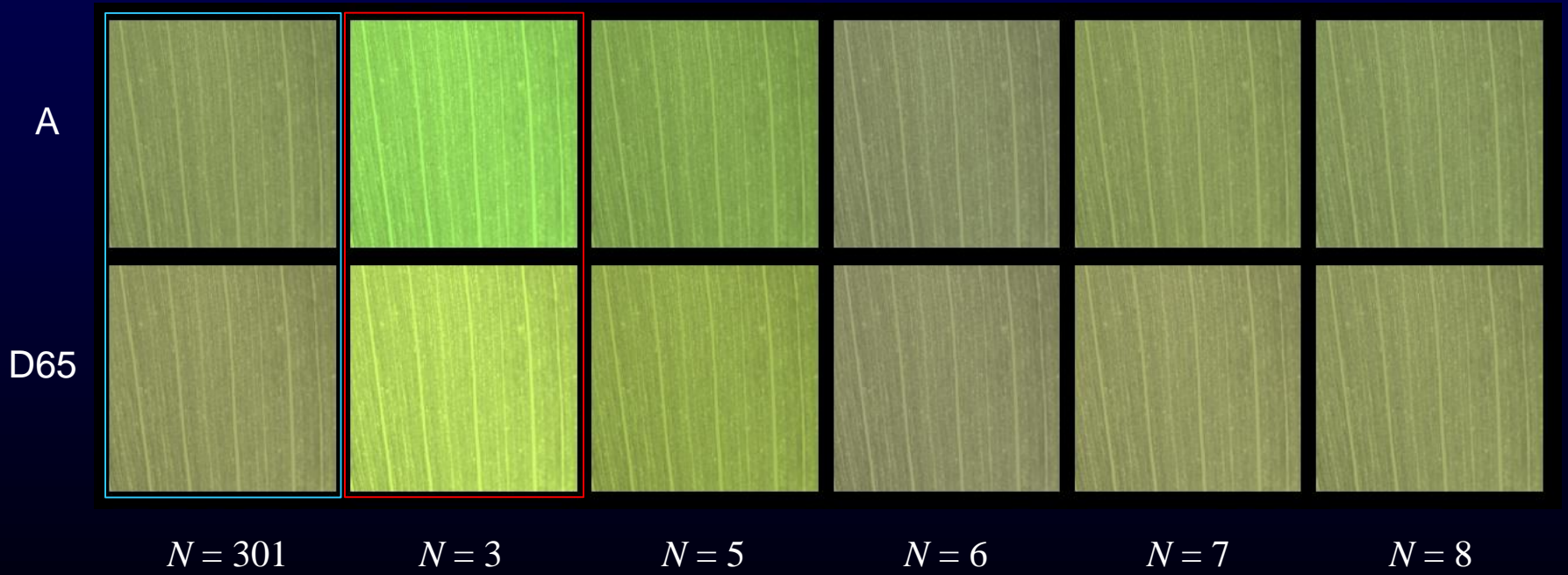
- Using 7 samples, the differences are below the perceptibility threshold (2.3)

- CIELAB differences

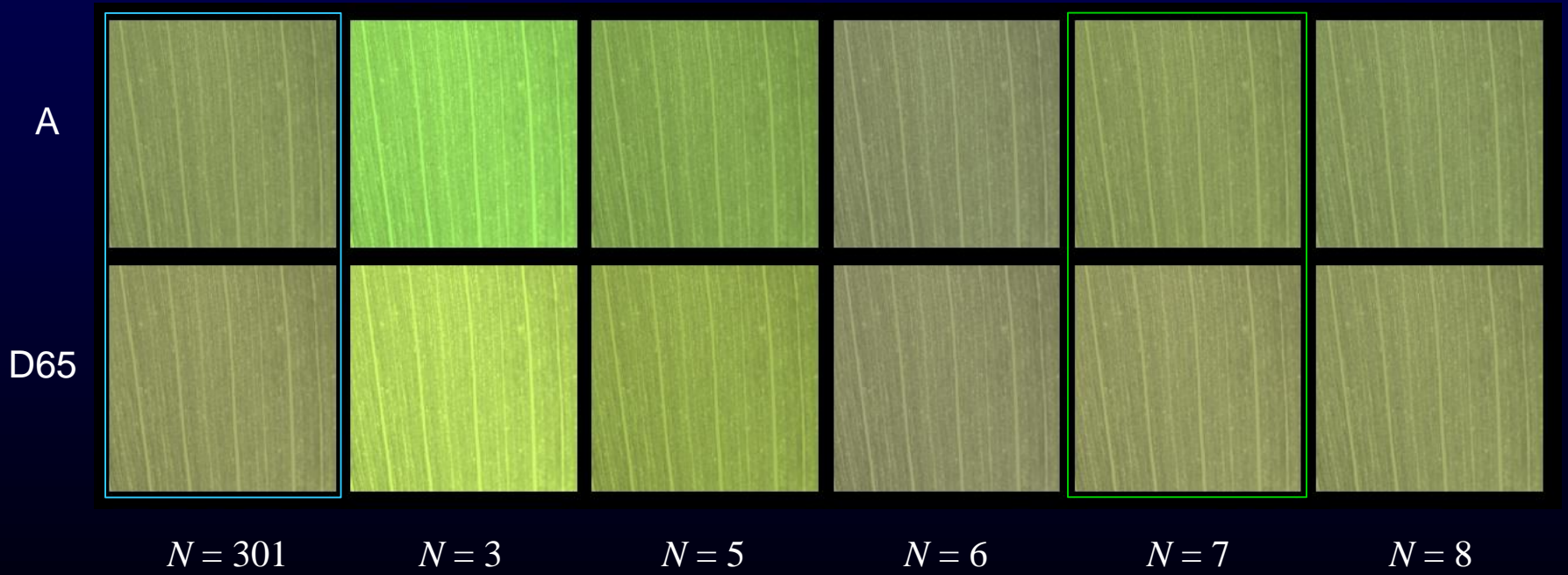
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- Using 7 samples, the differences are below the perceptibility threshold (2.3)
- Using 8 samples, they become markedly below the threshold

- Swatches depicting foliar aggregated reflective and transmissive behaviour



- Swatches depicting foliar aggregated reflective and transmissive behaviour



- CIELAB differences

Illuminant	Aggregated Reflective and Transmissive Behaviour				
	$N = 3$	$N = 5$	$N = 6$	$N = 7$	$N = 8$
A	45.8230	17.1292	10.3191	1.9361	1.1052
D65	46.1237	25.3037	9.8280	1.7179	0.8424

- Using 7 samples, the differences are below the threshold (2.3)

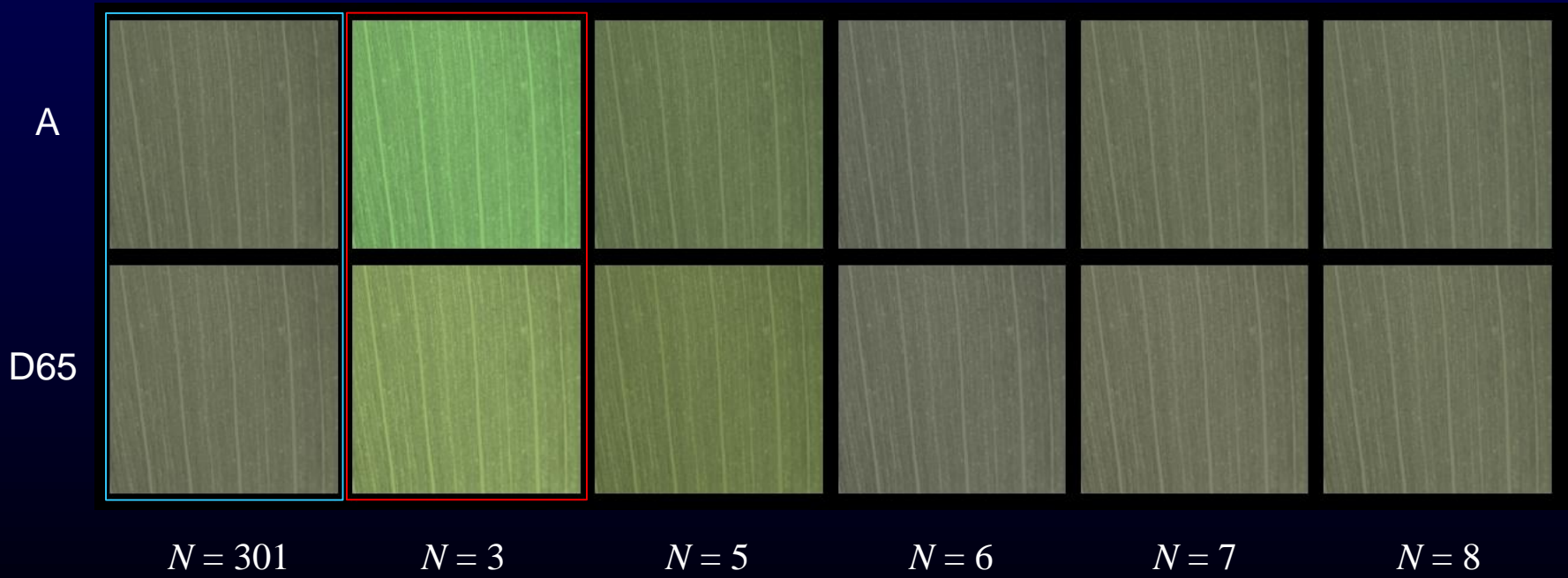
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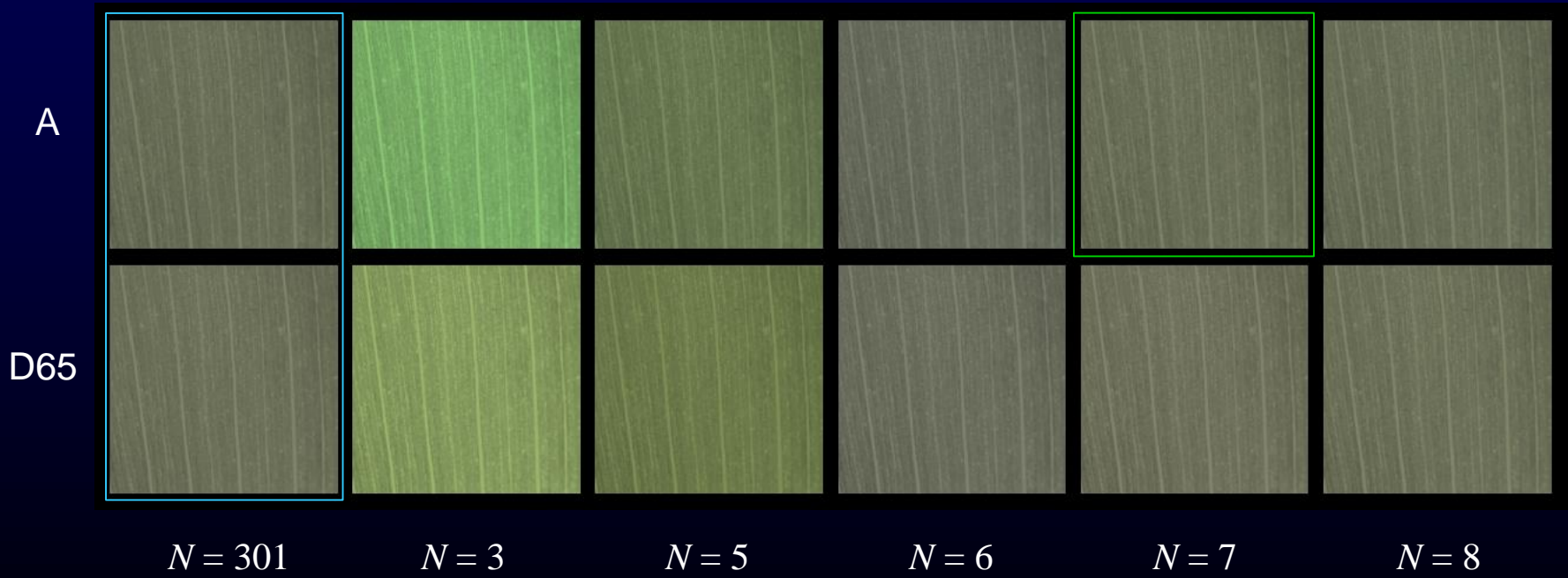
➤ Specimen M2

- Swatches depicting reflective behaviour



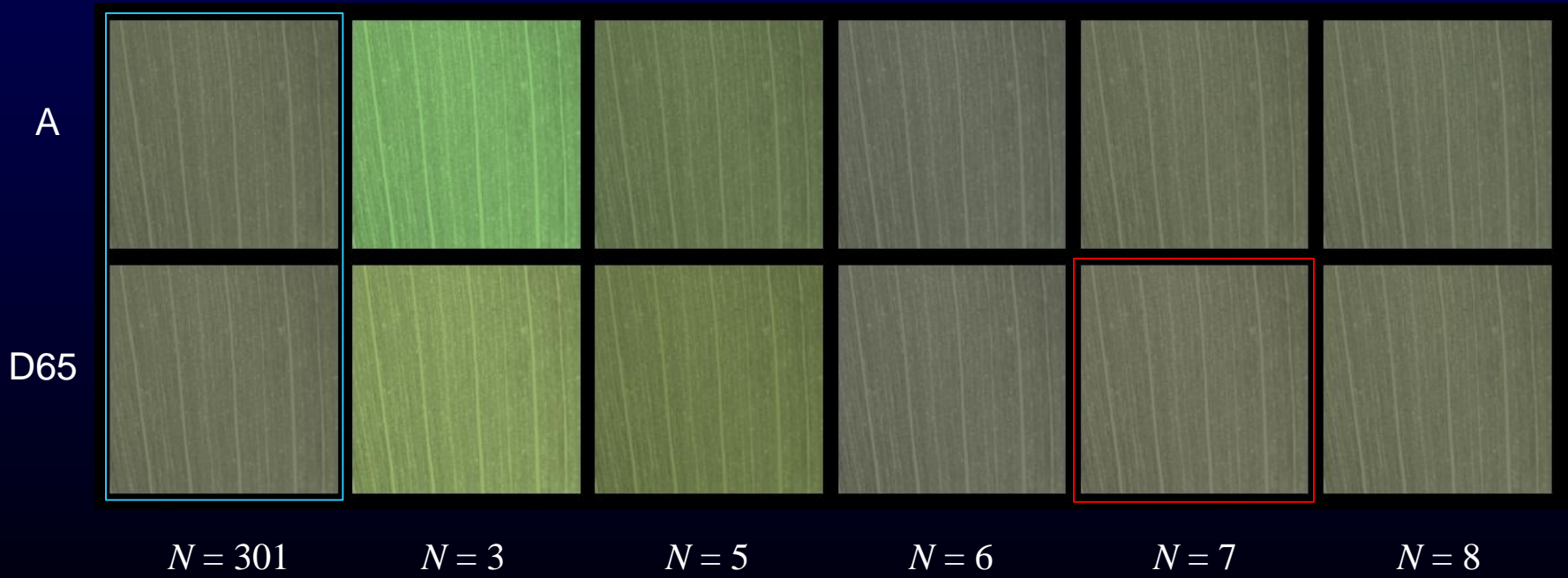
➤ Specimen M2

- Swatches depicting reflective behaviour



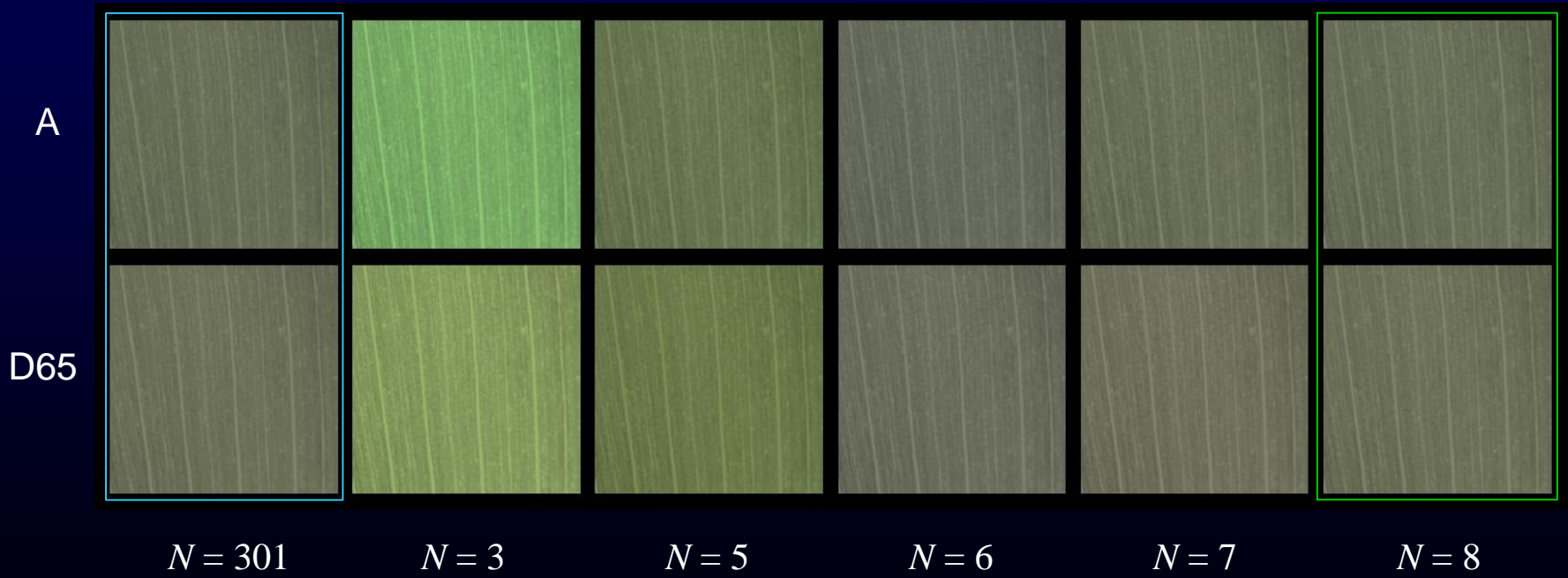
➤ Specimen M2

- Swatches depicting reflective behaviour



➤ Specimen M2

- Swatches depicting reflective behaviour



- CIELAB differences

Illuminant	Reflective Behaviour				
	$N = 3$	$N = 5$	$N = 6$	$N = 7$	$N = 8$
A	40.4986	9.7705	5.3314	0.6346	0.8277
D65	31.1737	18.0523	4.6922	2.7911	0.7095

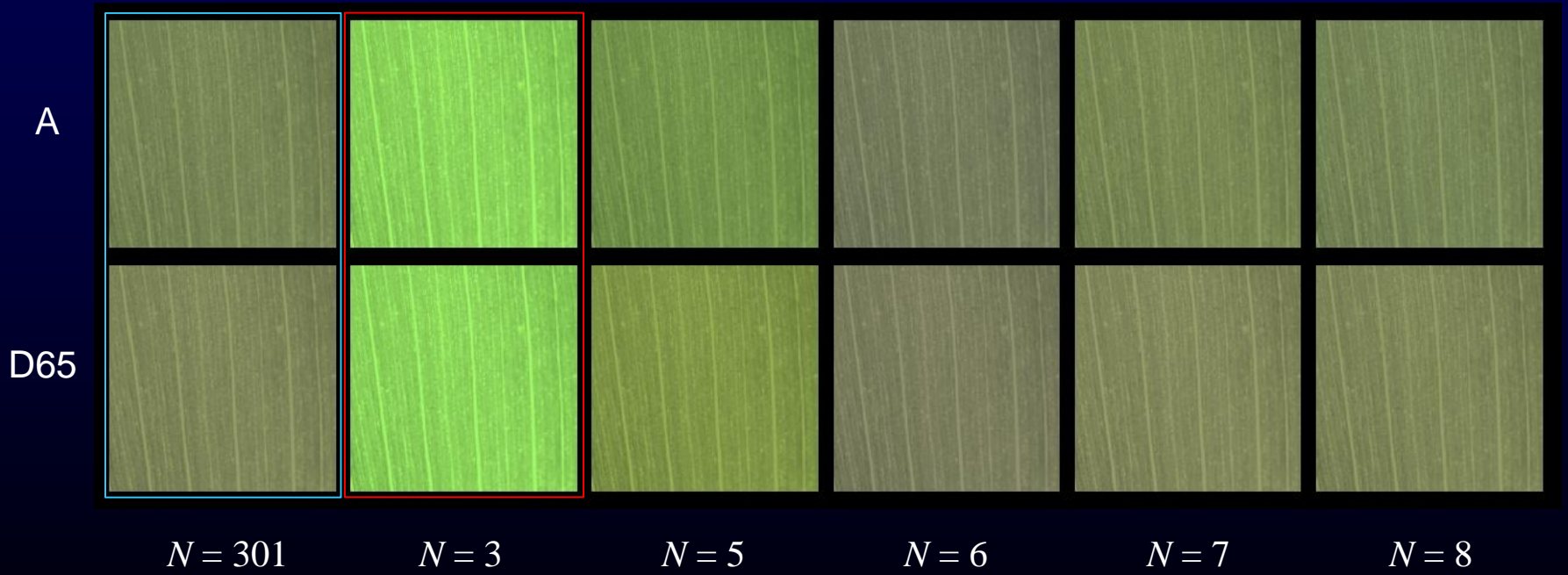
- Using 7 samples, **not** both differences are below the threshold (2.3)

- CIELAB differences

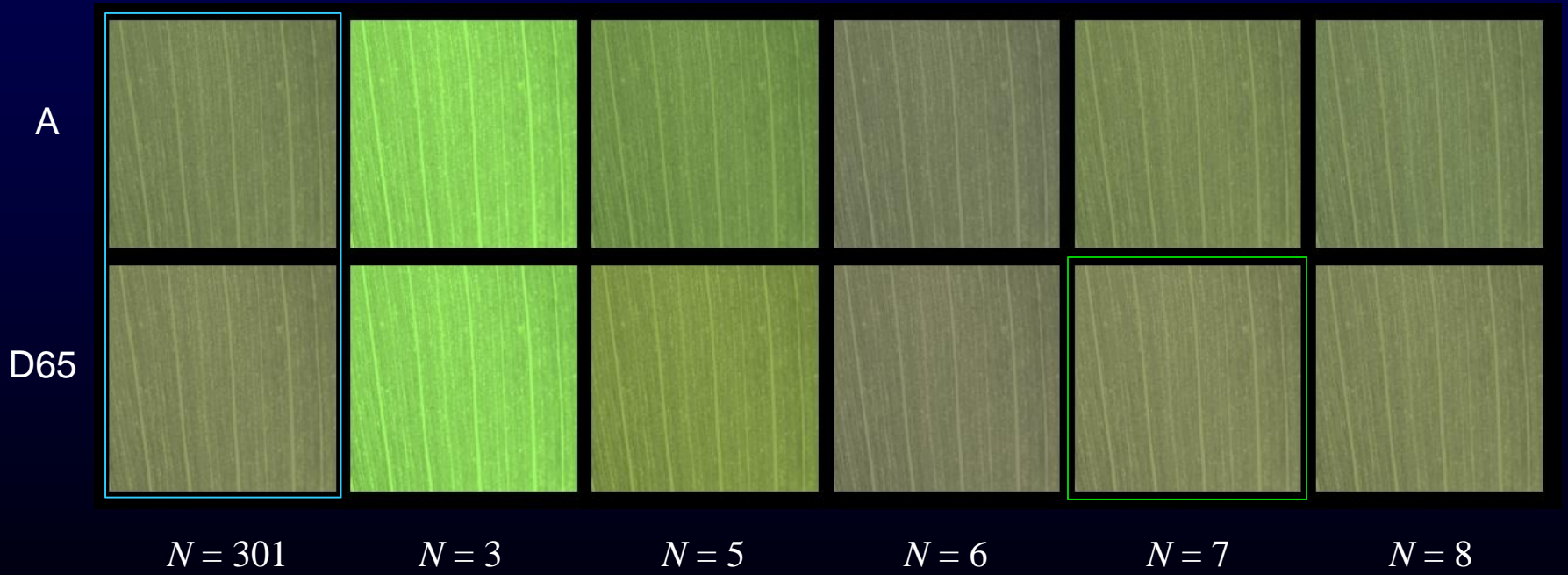
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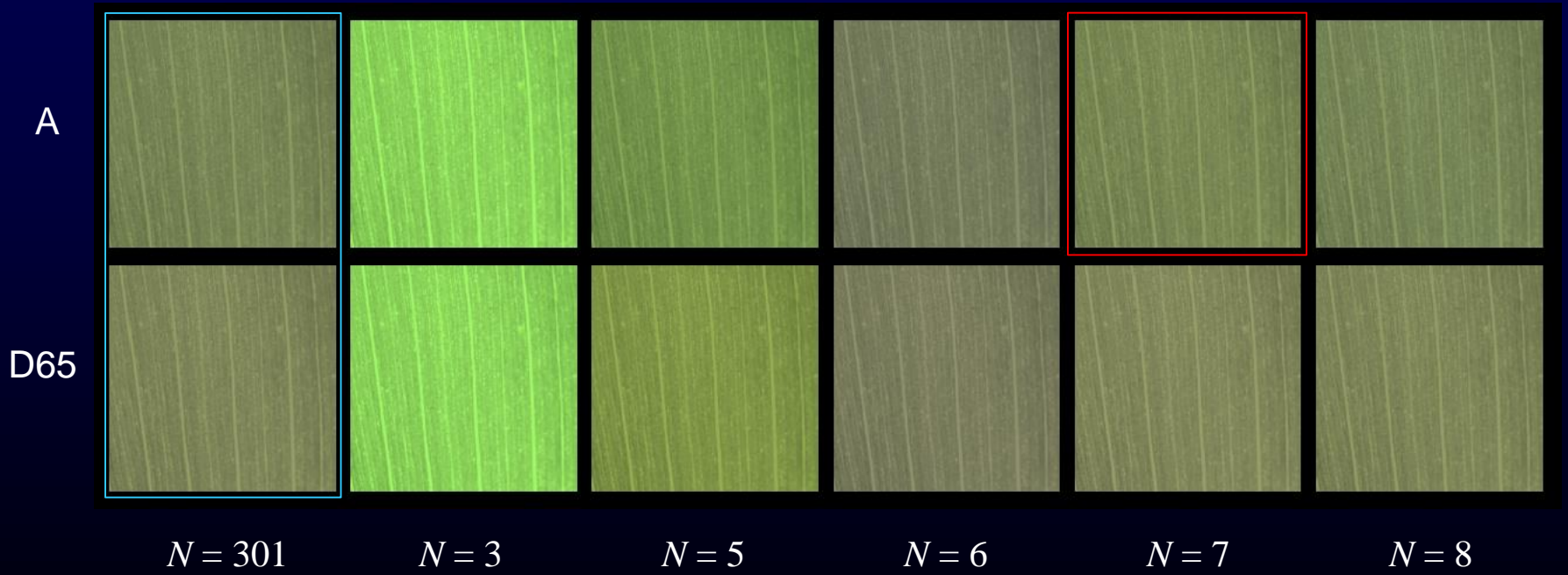
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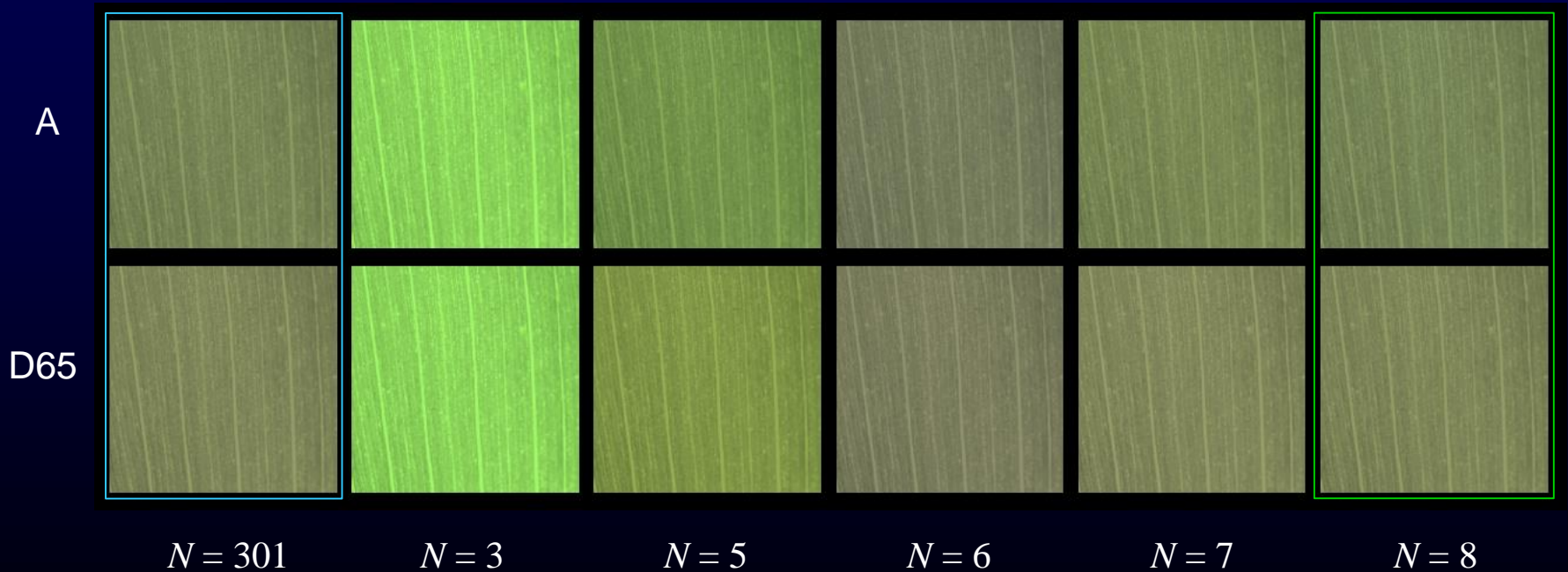
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- Swatches depicting foliar aggregated reflective and transmissive behaviour



- Swatches depicting foliar aggregated reflective and transmissive behaviour



- CIELAB differences

Illuminant	Aggregated Reflective and Transmissive Behaviour				
	$N = 3$	$N = 5$	$N = 6$	$N = 7$	$N = 8$
A	53.5902	17.3101	9.9848	2.4778	0.7864
D65	43.4365	24.4057	9.9055	1.4961	0.6757

- Using 7 samples, **not** both differences are below the threshold (2.3)

- CIELAB differences

Illuminant	Aggregated Reflective and Transmissive Behaviour				
	$N = 3$	$N = 5$	$N = 6$	$N = 7$	$N = 8$
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➤ Practical ramifications

- Our preliminary findings suggest that 8 spectral samples may be sufficient to obtain a high-fidelity reproduction of the colors of healthy maize leaves
- It may be unfeasible to find the exact number of samples that would work for:
 - all the different sampling schemes and distinct illumination/viewing geometries that could be employed in the monitoring of these plants
 - the relatively broad range of maize specimens' spectral signatures
- Nonetheless, our preliminary findings provide a basis for future experiments involving maize and other C4 species (*e.g.*, sugarcane)

Conclusion

➤ Recap

- Different approaches can be employed to monitor maize crops:
 - involving the calculation of spectral vegetation indices
 - involving the analysis of foliar chromatic attributes
- None can be considered the “magic bullet” capable of providing the “best” feedback for all instances
- All rely, directly or indirectly, on the interpretation of spectral signatures
- Hence, the need for more cost-effective strategies to sample those signatures

➤ Outlook

- The development of new technologies in this area will likely involve the implementation and combination of different methods
- It may be beneficial to design sensors to acquire a (low) number of spectral samples that can also be used in the creation of compact & high-fidelity leaf color databases
- These databases could potentially be extended to other C4 species (e.g., sugarcane) with similar characteristics



Thank you!

Questions?